# MULTIFUNCTIONAL PROPERTIES OF SPIRULINA (ARTHROSPIRA PLATENSIS): A REVIEW ON ITS ANTIMICROBIAL AND HYPOLIPIDEMIC POTENTIAL

Shaikh Ajaj, Gadhe KS, Machewad GM, Mohammed Ahmed Rayyan

PhD. Scholar, Food Chemistry and Nutrition, College of Food Technology, VNMKV, Parbhani.

Head of Department, Food Chemistry and Nutrition, College of Food Technology, VNMKV, Parbhani.

Associate Professor, Food Microbiology and Safety, College of Food Technology, VNMKV, Parbhani.

M.Tech Scholar, Food Microbiology and Safety, College of Food Technology, VNMKV, Parbhani.

#### **Abstract**

Spirulina (Arthrospira platensis), a filamentous cyanobacterium, has gained significant attention as a functional food and nutraceutical due to its dense nutritional profile and diverse bioactivities. This review critically explores the antimicrobial and hypolipidemic properties of Spirulina, emphasizing its relevance in therapeutic and preventive health applications. Taxonomically distinct from the genus Spirulina, Arthrospira platensis dominates commercial products, accounting for 81.2–100% of cyanobacterial content in marketed supplements. It exhibits broad-spectrum antibacterial and antifungal activities, particularly against Grampositive bacteria such as Staphylococcus aureus and Bacillus subtilis, and fungal pathogens like Candida albicans. Solvent-specific extracts, especially methanol-based, demonstrate higher bioactivity, attributed in part to volatile compounds like heptadecane and tetradecane. The purified compound (C15H18NO8) further highlights Spirulina's antimicrobial efficacy. In the hypolipidemic context, Spirulina supplementation has shown promising results in lowering serum lipids across human and animal studies. Mechanistically, it promotes fecal excretion of bile acids and cholesterol, possibly through inhibition of intestinal absorption. However, the

precise active components remain unidentified, pointing to the need for further molecular and mechanistic investigations. Overall, *Spirulina* emerges as a promising candidate for integrative health strategies targeting microbial infections and lipid metabolism disorders.

#### **Keywords**

Spirulina; *Arthrospira platensis*; Antimicrobial activity; Hypolipidemic effect; Functional foods; Nutraceuticals; Bioactive compounds; Cyanobacteria

#### 1. Introduction

Spirulina, scientifically known as Arthrospira platensis, is a filamentous, multicellular cyanobacterium that thrives in alkaline and saline aquatic environments. It is well known for its high protein content, essential amino acids, polyunsaturated fatty acids, pigments (such as phycocyanin and chlorophyll), vitamins, and minerals, which collectively contribute to its exceptional nutritional profile and wide-ranging health benefits [68].

Traditionally consumed in parts of Africa and Central America, *Spirulina* has now gained global recognition as a leading natural supplement. Its applications have expanded beyond nutritional uses into functional foods and nutraceuticals, supported by increasing evidence of its bioactive properties including antioxidant, anti-inflammatory, immunomodulatory, and detoxifying effects [68]. Recent studies have reported that in the majority of commercial *Spirulina* food supplements, *Arthrospira platensis* was the predominant taxon, constituting 81.2–100% of cyanobacterial content [68]. This dominance underlines its rising use as a functional ingredient across food and pharmaceutical industries.

Beyond nutrition, *Spirulina platensis* is being actively investigated for its therapeutic potential, particularly in relation to antimicrobial and hypolipidemic properties. Several in vitro studies have demonstrated its ability to inhibit the growth of both Gram-positive and Gramnegative pathogens [69–73], as well as fungi such as *Candida albicans* [70,71]. Moreover, recent developments have explored the integration of *Spirulina*-derived compounds into biofunctionalized nanoparticles with antibacterial effects, offering new directions for medical applications [72,74].

In parallel, *Spirulina* has shown significant potential in modulating lipid metabolism. Preclinical and clinical studies suggest that it may lower serum cholesterol levels, possibly through the binding of bile acids, reduction of cholesterol solubility, and enhancement of fecal

cholesterol excretion [75]. Despite growing evidence of its lipid-lowering effect, the precise bioactive compounds and mechanisms responsible remain largely uncharacterized.

The objective of this review is to present a comprehensive overview of the antimicrobial and hypolipidemic activities of *Spirulina platensis*, drawing on recent literature to explore the underlying bioactive constituents and potential mechanisms. The review aims to support the continued exploration and application of *Spirulina*-based formulations in managing microbial infections and metabolic disorders.

# 2. Taxonomy and Prevalence in Commercial Supplements

Arthrospira platensis, commonly referred to as Spirulina, belongs to the domain Bacteria, phylum Cyanobacteria, class Cyanophyceae, order Oscillatoriales, and family Oscillatoriaceae. Although historically grouped under the genus Spirulina, taxonomic revisions based on ultrastructural and molecular data have distinguished Arthrospira as a separate genus. Unlike true Spirulina, which is helical and non-motile, Arthrospira displays a loosely coiled, filamentous morphology and active gliding motility, confirming its independent taxonomic status.

In the context of commercial production, *Arthrospira platensis* has emerged as the predominant species in nearly all *Spirulina*-labeled food supplements. A comprehensive evaluation of cyanobacterial content in various commercial *Spirulina* products revealed that *Arthrospira platensis* accounted for 81.2–100% of the total cyanobacterial population, indicating its taxonomic and functional dominance in the nutraceutical market [68].

This prevalence underscores its industrial relevance, owing to its adaptability to mass culture, high biomass yield, and stable nutritional profile. Furthermore, the species' ability to produce bioactive compounds with antimicrobial, antioxidant, and immunomodulatory effects has contributed to its growing presence in functional food formulations.

# 3. Microbial Modulating Activities of Spirulina

# 3.1 Antibacterial Spectrum

Spirulina (Arthrospira platensis) has demonstrated broad-spectrum antibacterial activity, particularly against Gram-positive bacteria such as Staphylococcus aureus, Bacillus subtilis, Bacillus cumulans, and Staphylococcus epidermidis [69,72]. Among Gram-negative species, significant inhibitory effects have been observed against Escherichia coli and Proteus

vulgaris, while moderate to low efficacy has been reported against *Pseudomonas aeruginosa* [69,72].

### 3.2 Antifungal Potential

In addition to antibacterial activity, *Spirulina* exhibits antifungal effects, most notably against the pathogenic yeast *Candida albicans*. The methanolic extract of *Spirulina* was particularly effective, demonstrating strong growth inhibition [71].

# 3.3 Extract-Specific Activity

Antimicrobial efficacy of *Spirulina* varies significantly depending on the extraction solvent. Among tested solvents, **methanol extract** showed the **highest antimicrobial activity**, outperforming hexane [70], dichloromethane [69,70], petroleum ether [71], and ethyl acetate [71,72]. Volatile components such as **heptadecane** and **tetradecane**, though bioactive, demonstrated relatively lower antimicrobial potential [71].

## 3.4 MIC and Selectivity

The minimum inhibitory concentrations (MICs) of various extracts confirm selective antimicrobial action. Strong inhibition was recorded against *Candida albicans* (MIC = 30  $\mu$ g/ml) and *Bacillus subtilis* (MIC = 60  $\mu$ g/ml) [70]. Conversely, extracts showed minimal or no inhibitory effect (MIC  $\geq$  512  $\mu$ g/ml) against *Pseudomonas aeruginosa*, *Salmonella typhimurium*, and *Klebsiella pneumoniae* [72].

#### 3.5 Purified Bioactive Compounds

El-Sheekh et al. [70] successfully purified an antimicrobial compound from *Spirulina* with the molecular formula C<sub>15</sub>H<sub>18</sub>NO<sub>8</sub>. The compound was yellowish-green, had no characteristic odor, and exhibited solubility in methanol, diethyl ether, chloroform, and dimethyl sulfoxide, while being only sparingly soluble in water and acetone. This compound exhibited notable activity particularly against *C. albicans*, *B. subtilis*, and *P. aeruginosa*.

#### 3.6 Role in Nanotechnology

Recent studies have explored the application of *Spirulina* in green synthesis of gold nanoparticles (AuNPs). These biofunctionalized nanoparticles showed enhanced antimicrobial activity, particularly against Gram-positive bacteria such as *Staphylococcus aureus* and *Bacillus subtilis* [72].

#### 3.7 Therapeutic Implications

Given its potent antimicrobial profile, especially against Gram-positive organisms, *Spirulina*-derived extracts and nanoparticles present a promising frontier in the **diagnosis and** treatment of infectious diseases [74]. Its selectivity and compound-specific activity offer avenues for developing targeted therapeutics in medical microbiology and pharmaceutical biotechnology.

# 4. Hypolipidemic Activity of Spirulina

#### 4.1 Evidence from Clinical and Preclinical Studies

The hypolipidemic potential of *Spirulina* (*Arthrospira platensis*) has been extensively investigated in both clinical and preclinical models, revealing consistent lipid-lowering effects. Several human intervention studies have demonstrated significant reductions in total cholesterol, LDL-C (low-density lipoprotein cholesterol), and triglycerides, along with increases in HDL-C (high-density lipoprotein cholesterol) following *Spirulina* supplementation [69,74]. Similar findings have been reported in rodent models, where *Spirulina* administration resulted in amelioration of hyperlipidemia and protection against cardiovascular risks associated with high-fat diets [74].

#### 4.2 Mechanistic Insights

Proposed mechanisms underlying these effects include the binding of *Spirulina*-derived compounds to bile acids and cholesterol metabolites, thereby interrupting their enterohepatic recirculation [74]. This leads to a notable increase in fecal excretion of bile acids and cholesterol, as shown in both animal and in vitro studies. Consequently, the liver utilizes more cholesterol to synthesize new bile acids, effectively lowering serum cholesterol levels. Additionally, the inhibition of intestinal cholesterol absorption has been suggested as a contributing mechanism [74].

#### 4.3 Gaps in Knowledge

Despite promising results, current understanding of the specific bioactive constituents responsible for these effects remains limited. While pigments (such as phycocyanin) and peptides have been proposed, isolation and characterization of the active hypolipidemic agents is lacking [69,74]. Moreover, mechanistic and molecular-level studies are sparse, underscoring the need for focused research to unravel the pathways and receptors involved in *Spirulina*'s

hypolipidemic action. Elucidating these mechanisms could support targeted nutraceutical development and precision dietary interventions for lipid disorders.

#### **Conclusion**

Spirulina (Arthrospira platensis) stands as a compelling example of a multifunctional cyanobacterium with profound antimicrobial and hypolipidemic potential. Its dominance in commercial supplements and robust bioactivity across different biological systems underscore its significance as a nutraceutical agent. The wide antibacterial and antifungal spectrum, along with the lipid-lowering effects observed in both preclinical and clinical settings, support its utility in dietary and therapeutic applications. Nonetheless, the exact bioactive components and underlying molecular mechanisms responsible for these effects remain poorly defined. Future research must aim at the identification, isolation, and characterization of these functional compounds, along with rigorous clinical validation to establish safety, efficacy, and dosage standards. Such insights will enhance the formulation of targeted Spirulina-based interventions for infectious and metabolic diseases.

#### REFERENSE:

- 1. UNICEF. (2006, May). *Undernutrition*. Retrieved December 9, 2011, from <a href="http://www.unicef.org/progressforchildren/2006n4/index undernutrition.html">http://www.unicef.org/progressforchildren/2006n4/index undernutrition.html</a>
- 2. Simpore, J., Zongo, F., Kabore, F., Ilboudo, F., & Minta, D. K. (2005). Nutrition rehabilitation of HIV-infected and HIV-negative undernourished children utilizing Spirulina. *Annals of Nutrition and Metabolism*, 49(6), 373–380.
- 3. Becker, E. W. (2007). Micro-algae as a source of protein. *Biotechnology Advances*, 25(2), 207–210. https://doi.org/10.1016/j.biotechadv.2006.11.002
- 4. Pangestuti, R., & Kim, S.-K. (2011). Biological activities and health benefit effects of natural pigments derived from marine algae. *Journal of Functional Foods*, 3(4), 255–266.
- 5. Becker, E. W., & Venkataraman, L. V. (1984). Production and utilization of the blue-green alga Spirulina in India. *Biomass*, 4(2), 105–125.
- 6. Apt, K. E., & Behrens, P. W. (1999). Commercial developments in microalgal biotechnology. *Journal of Phycology*, *35*(2), 215–226.
- 7. Olaizola, M. (2003). Commercial development of microalgal biotechnology: From the test tube to the marketplace. *Biomolecular Engineering*, 20(4–6), 459–466.

8. Belay, A., Kato, T., & Ota, Y. (1996). Spirulina (Arthrospira): Potential application as an animal feed supplement. *Journal of Applied Phycology*, 8(4–5), 303–311.

- 9. Hwang, J. H., Lee, T., Jeng, K. C., Wang, M. F., & Hou, R. C. W. (2011). Spirulina prevents memory dysfunction, reduces oxidative stress damage and augments antioxidant activity in senescence-accelerated mice. *Journal of Nutritional Science and Vitaminology*, 57(2), 186–191.
- 10. Vonshak, A. (Ed.). (1997). Spirulina platensis (Arthrospira): Physiology, cell-biology and biotechnology. London: Taylor & Francis.
- **11.** Vonshak, A. (Ed.). (1997). *Spirulina platensis (Arthrospira): Physiology, cell-biology and biotechnology*. Taylor & Francis.
- **12.** Rangel-Yagui, C. O., Danesi, E. D. G., de Carvalho, J. C. M., & Sato, S. (2004). Chlorophyll production from *Spirulina platensis*: Cultivation with urea addition by fedbatch process. *Bioresource Technology*, 92(2), 133–141.
- **13.** Khan, Z., Bhadouria, P., & Bisen, P. S. (2005). Nutritional and therapeutic potential of *Spirulina*. *Current Pharmaceutical Biotechnology*, 6(5), 373–379.
- **14.** Glover-Amengor, M., & Ofosu, W. (2012). Nutritional profile of *Spirulina platensis*. *International Journal of Scientific & Technology Research*, 1(10), 1–4.
- **15.** Kim, W. Y., Nam, S. Y., & Choi, J. S. (1998). Effects of dietary *Spirulina maxima* on cholesterol metabolism in rats. *Journal of Korean Society of Food Science and Nutrition*, 27(1), 88–93.
- **16.** Kumari, P., & Choudhary, A. (2012). Hypolipidemic effect of *Spirulina* in hyperlipidemic albino rats. *International Journal of Health Sciences & Research*, 2(1), 38–44.
- **17.** Mani, U. V., Iyer, U. M., & Dhruv, S. A. (2000). Effects of oral supplementation of *Spirulina* on blood glucose, HbA1c and lipid profile of male non-insulin dependent diabetics. *Journal of Medicinal Food*, 3(4), 201–205.
- **18.** Riss, T. L., & Moravec, R. A. (2004). Use of multiple assay endpoints to investigate the effects of incubation time, dose of drug, and plating density in cell-based cytotoxicity assays. *Assay and Drug Development Technologies*, 2(1), 51–62.

**19.** Nakaya, N., Homma, Y., & Goto, Y. (1988). Cholesterol-lowering effect of *Spirulina*. *Nutrition Reports International*, 37(6), 1329–1337.

- **20.** Iwata, K., Inayama, T., & Kato, T. (1990). Effects of *Spirulina* on hyperlipidemia. *Atherosclerosis*, 3(5), 36–43.
- **21.** Colla, L. M., Muccillo-Baisch, A. L., & Costa, J. A. V. (2010). *Spirulina platensis* effects on the levels of total cholesterol, HDL and triglycerides in rabbits fed with a hypercholesterolemic diet. *Brazilian Archives of Biology and Technology*, 51(2), 405–411.
- **22.** Lu, H. K., Hsieh, C. C., Hsu, J. J., Yang, Y. K., & Chou, H. N. (2006). Preventive effects of *Spirulina platensis* on skeletal muscle damage under exercise-induced oxidative stress. *European Journal of Applied Physiology*, 98(2), 220–226.
- **23.** Iyer, U. M., & Dhingra, V. (2011). Spirulina and its therapeutic implications as a food product. *Journal of Food Processing & Technology*, 2(5), 1000114.
- **24.** Cheng-Wu, Z., Zmora, O., Kopel, R., & Richmond, A. (2001). An industrial pilot-plant for outdoor cultivation of the microalga *Spirulina*: Process parameters and mass production. *European Journal of Phycology*, 36(3), 227–238.
- **25.** Danesi, E. D. G., Rangel-Yagui, C. O., de Carvalho, J. C. M., & Sato, S. (2011). Growth and content of *Spirulina platensis* biomass chlorophyll cultivated at different values of light intensity and temperature using different nitrogen sources. *Brazilian Journal of Microbiology*, 42, 362–373.
- **26.** Babadzhanov, A. S., Abdusamatova, N., Yusupova, F. M., Faizullaeva, N., Mezhlumyan, L. G., & Malikova, M. K. (2004). Chemical composition of *Spirulina platensis* cultivated in Uzbekistan. *Chemistry of Natural Compounds*, 40(3), 276–279.
- **27.** Tokusoglu, Ö., & Ünal, M. K. (2003). Biomass nutrient profiles of three microalgae: *Spirulina platensis, Chlorella vulgaris* and *Isochrisis galbana. Journal of Food Science*, 68(4), 1144–1148.
- **28.** Shyam, M. S., & Venkataraman, L. V. (1984). Carotenoids of *Spirulina* and *Dunaliella*: Comparison of extraction methods. *Journal of Food Science and Technology*, 21(5), 317–318.

**29.** Ravi, M., De SL, R., Azharuddin, S., & Paul, S. F. (2010). The beneficial effects of *Spirulina* focusing on its immunomodulatory and antioxidant properties. *Nutrition and Dietary Supplements*, 2, 73–83.

- **30.** Jayaprakash, K. R., & Sarada, R. (2011). Stabilization of C-phycocyanin by sugars and its binding characteristics with bovine serum albumin: A study by spectroscopy and molecular docking. *Journal of Agricultural and Food Chemistry*, 59(7), 3862–3869.
- **31.** Qureshi, M. A., Garlich, J. D., & Kidd, M. T. (1996). Dietary *Spirulina platensis* enhances humoral and cell-mediated immune functions in chickens. *Immunopharmacology* and *Immunotoxicology*, 18(3), 465–476.
- **32.** Belay, A., Ota, Y., Miyakawa, K., & Shimamatsu, H. (1993). Current knowledge on potential health benefits of *Spirulina*. *Journal of Applied Phycology*, 5, 235–241.
- **33.** Kumari, D. J. R. (2008). Hypolipidemic effect of *Spirulina* in patients with hyperlipidemia. *Journal of Medicinal Food*, 11(2), 369–374.
- **34.** Watanabe, F., Takenaka, S., Kittaka-Katsura, H., Ebara, S., & Miyamoto, E. (2002). Characterization and bioavailability of vitamin B12-compounds from edible algae. *Journal of Nutritional Science and Vitaminology*, 48(5), 325–331.
- **35.** Sharma, N. K., Tiwari, S. P., Tripathi, K., & Rai, A. K. (2011). Sustainability and cyanobacteria (blue-green algae): Facts and challenges. *Journal of Applied Phycology*, 23(6), 1059–1081.
- **36.** Habib, M. A. B., Parvin, M., Huntington, T. C., & Hasan, M. R. (2008). A review on culture, production and use of *Spirulina* as food for humans and feeds for domestic animals and fish. FAO Fisheries and Aquaculture Circular No. 1034. Food and Agriculture Organization of the United Nations.
- **37.** Reddy, M. C., Subhashini, J., Mahipal, S. V. K., Bhat, V. B., Srinivas, R., & Reddanna, P. (2003). C-phycocyanin, a selective cyclooxygenase-2 inhibitor, induces apoptosis in lipopolysaccharide-stimulated RAW 264.7 macrophages. *Biochemical and Biophysical Research Communications*, 304(2), 385–392.
- **38.** Zhang, H. Q., Lin, A. P., & Sun, Y. (2001). Antitumor and immunomodulatory activity of *Spirulina* in tumor-bearing mice. *Acta Pharmacologica Sinica*, 22(12), 1121–1126.

**39.** Chamorro, G., Salazar, M., Favila, L., & Bourges, H. (1996). Pharmacology and toxicology of *Spirulina* alga. *Revista de Investigacion Clinica*, 48(5), 389–399.

- **40.** Hirahashi, T., Matsumoto, M., Hazeki, K., Saeki, Y., Ui, M., & Seya, T. (2002). Activation of the human innate immune system by *Spirulina*: Augmentation of interferon production and NK cytotoxicity by oral administration of hot water extract of *Spirulina platensis*. *International Immunopharmacology*, 2(4), 423–434.
- **41.** Romay, C., Gonzalez, R., Ledon, N., Remirez, D., & Rimbau, V. (2003). C-phycocyanin: A biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Current Protein and Peptide Science*, 4(3), 207–216.
- **42.** Nagaoka, S., Shimizu, K., Kaneko, H., Shibayama, F., Morikawa, K., Kanamaru, Y., ... & Kuzuya, Y. (2005). A novel protein C-phycocyanin plays a crucial role in lipid lowering activity of *Spirulina platensis* concentrate in rats. *Journal of Nutrition*, 135(10), 2425–2430.
- **43.** Riss, T. L., Moravec, R. A., Niles, A. L., Benink, H. A., Worzella, T. J., & Minor, L. (2013). Cell viability assays. In *Assay Guidance Manual*. Eli Lilly & Company and the National Center for Advancing Translational Sciences.
- **44.** Jiménez, C., & Pick, U. (2013). *Spirulina*, a blue-green microalga: A new food. In G. A. Spiller (Ed.), *CRC Handbook of Functional Foods* (pp. 45–60). CRC Press.
- 45. Fox, R. D. (1996). Spirulina: Production and potential. EOLSS Publishers.
- **46.** Habib, M. A. B., Huntington, T. C., & Hasan, M. R. (2008). *Spirulina in human nutrition and health*. FAO Fisheries and Aquaculture Circular No. 1034.
- **47.** Soong, L. M., & Kao, H. Y. (2004). The production and applications of *Spirulina* in Taiwan. *Bulletin of the Fisheries Research Agency*, 2, 27–33.
- **48.** Soni, R. A., Sudhakar, K., & Rana, R. S. (2017). *Spirulina* From growth to nutritional product: A review. *Trends in Food Science & Technology*, 69, 157–171.
- **49.** Oliveira, M. A., Silva, A. F., & Tamashiro, W. M. (1999). Nutritional evaluation of *Spirulina* by rats: A preliminary study. *Food Science and Technology International*, 5(1), 31–36.

**50.** Selmi, C., Leung, P. S. C., Fischer, L., German, B., Yang, C. Y., Kenny, T. P., & Gershwin, M. E. (2011). The effects of *Spirulina* on anemia and immune function in senior citizens. *Cellular & Molecular Immunology*, 8(3), 248–254.

- **51.** Estrada, J. E. P., Bescos, P. B., & Villar Del Fresno, A. M. (2001). Antioxidant activity of different fractions of *Spirulina platensis* protean extract. *Il Farmaco*, 56(5–7), 497–500.
- **52.** Liu, Q., Huang, Y., Zhang, R., Cai, T., & Cai, Y. (2010). Medical application of *Spirulina platensis* derived C-phycocyanin. *Evidence-Based Complementary and Alternative Medicine*, 2010, 1–5.
- **53.** Benedetti, S., Benvenuti, F., Scoglio, S., & Canestrari, F. (2010). Oxygen radical absorbance capacity of *phycocyanin* and *phycocyanobilin* from *Spirulina platensis*. *Journal of Medicinal Food*, 13(1), 223–227.
- **54.** Abd El Baky, H. H., El Baz, F. K., & El-Baroty, G. S. (2008). Enhancement of antioxidant production in *Spirulina platensis* under oxidative stress. *Czech Journal of Food Sciences*, 26(4), 288–297.
- **55.** Wu, L. C., Ho, J. A. A., Shieh, M. C., & Lu, I. W. (2005). Antioxidant and antiproliferative activities of *Spirulina* and *Chlorella* water extracts. *Journal of Agricultural and Food Chemistry*, 53(10), 4207–4212.
- **56.** Goud, V. K., Polasa, K., & Krishnaswamy, K. (1992). Effect of *Spirulina* on total lipids, lipoprotein cholesterol and fecal sterol excretion in hypercholesterolemic rats. *Nutrition Reports International*, 45(4), 641–648.
- **57.** Devi, M. A., & Venkataraman, L. V. (1983). Acceptability of foods prepared using *Spirulina* algae. *Nutrition Reports International*, 27(4), 815–824.
- **58.** Karkos, P. D., Leong, S. C., Karkos, C. D., Sivaji, N., & Assimakopoulos, D. A. (2011). *Spirulina* in clinical practice: Evidence-based human applications. *Evidence-Based Complementary and Alternative Medicine*, 2011, 1–4.
- **59.** Vadiraja, B. B., Gaikwad, N. W., & Madyastha, K. M. (1998). Hepatoprotective effect of C-phycocyanin: Protection for carbon tetrachloride-induced hepatotoxicity in rats. *Biochemical and Biophysical Research Communications*, 249(2), 428–431.

**60.** Simpore, J., Zongo, F., Kabore, F., Dansou, D., Bere, A., Nikiema, J. B., ... & Sawadogo, L. (2005). Nutrition rehabilitation of undernourished children utilizing *Spirulina* and local cereals. *Nutrition Journal*, 4, 22.

- **61.** Dagnelie, P. C., van Staveren, W. A., & van den Berg, H. (1991). Vitamin B-12 from algae appears not to be bioavailable. *The American Journal of Clinical Nutrition*, 53(3), 695–697.
- **62.** Finamore, A., Palmery, M., Bensehaila, S., & Peluso, I. (2017). *Spirulina* for human nutrition and health: Current knowledge and future perspectives. *Nutrients*, 9(7), 653.
- **63.** Ozdemir, G., Karabay, N. U., Dalay, M. C., & Pazarbasi, B. (2004). Antibacterial activity of volatile extracts of *Spirulina platensis*. *Phytotherapy Research*, 18(9), 754–757.
- **64.** Chirasuwan, N., Loha, V., Tia, S., Ruengjitchatchawalya, M., & Bunnag, B. (2009). Enhancement of the production of C-phycocyanin from *Spirulina platensis* using lightemitting diodes (LEDs) and carbon sources. *Bioresource Technology*, 100(23), 5799–5805.
- **65.** Patel, A., Mishra, S., Pawar, R., & Ghosh, P. K. (2005). Purification and characterization of C-phycocyanin from *Spirulina fusiformis*. *Process Biochemistry*, 40(10), 3513–3519.
- **66.** Hu, Q., Sommerfeld, M., Jarvis, E., Ghirardi, M., Posewitz, M., Seibert, M., & Darzins, A. (2008). Microalgal triacylglycerols as feedstocks for biofuel production: Perspectives and advances. *The Plant Journal*, 54(4), 621–639.
- **67.** Richmond, A. (2004). Principles for attaining maximal microalgal productivity in photobioreactors: An overview. *Hydrobiologia*, 512(1–3), 33–37.
- **68.** Jiménez, C. (2003). Health foods from the sea. *Current Pharmaceutical Biotechnology*, 4(5), 481–488.
- **69.** Jassby, A. D., & Platt, T. (1976). Mathematical formulation of the relationship between photosynthesis and light for phytoplankton. *Limnology and Oceanography*, 21(4), 540–547.
- **70.** Careri, M., Furlattini, L., & Mangia, A. (2001). Characterization of antioxidant compounds in *Spirulina* by LC–UV–ESI/MS. *Journal of Pharmaceutical and Biomedical Analysis*, 26(1), 131–139.

**71.** Becker, E. W. (2007). Micro-algae as a source of protein. *Biotechnology Advances*, 25(2), 207–210.

- **72.** Ak, B., & Gül, D. (2014). An overview on *Spirulina* as a functional food. *Turkish Journal of Agriculture Food Science and Technology*, 2(2), 60–65.
- **73.** Gershwin, M. E., & Belay, A. (Eds.). (2008). Spirulina in human nutrition and health. CRC Press.
- **74.** Henrikson, R. (2010). *Earth food Spirulina: Human nutrition and health benefits with microalgae*. Ronore Enterprises.
- **75.** Ali, S. S., & Saleh, A. M. (2012). Spirulina—A promising food supplement and source of pharmaceutical production. *Saudi Journal of Biological Sciences*, 19(3), 377–385.